# Development of a PZA Pipeline to Improve Phenotypic and Molecular-Based Susceptibility Testing for PZA

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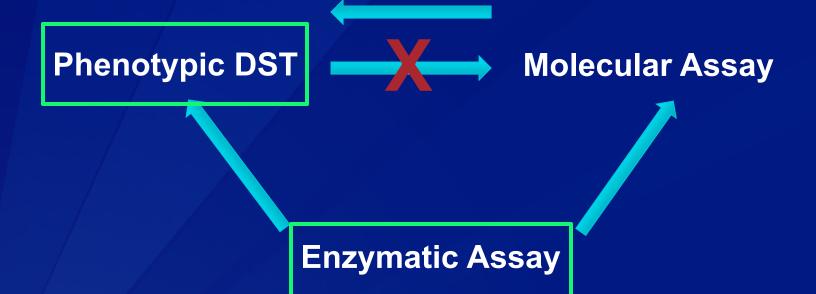
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National Center for HIV/AIDS, Viral Hepatitis, STD, and TB Prevention

Division of Tuberculosis Elimination







# PZA in vitro Activity

- Requires acidic culture conditions
  - Narrow pH range
    - Too alkaline = no PZA activity
    - Too acidic = inhibition of growth in drug-free media
    - pH 5.6 6.0 best
- Active against M. tuberculosis complex
  - Except M. bovis
    - Inherently PZA-resistant

# Phenotypic Detection of PZA Resistance

#### Indirect

- Pzase activity<sup>1</sup>
  - PZase positive = PZA-susceptible
  - PZase negative = PZA-resistant

#### Direct

- Culture methods
  - Growth on or in media containing PZA
  - Problematic
    - Both resistant and susceptible bacilli inhibited by acid pH
    - Inoculum size
  - Poor reproducibility<sup>2, 3, 4</sup>
    - False resistance and susceptibility
- 1. Wayne(1974), Amer. Rev. Resp. Dis., 109:147-151.
- 2. Chedore, et. al., (2010) J. Clin. Mirobiol,, 48:300-301
- 3. Chang, Yew and Zhang, (2011), Antimicrob. Agents & Chemo., 55:4499-4505.
- 4. Simons, et. al., (2012), J. Clin. Microbiol., 50:428-434.

# Factors Affecting in vitro PZA Activity

## pH of the medium<sup>1</sup>

- Can inhibit growth (~ 10% of strains)
- Theoretical calculations<sup>1</sup> (Henderson-Hasselbach equation)
  - If PZA MIC = 50 μg/ml @ pH 5.5
    - $_{\circ}$  MIC @ pH 6.1 = 200  $\mu$ g/ml
- Experimental results correlate well<sup>2</sup>

#### Inoculum size<sup>1</sup>

- Large inoculum reduces PZA activity
  - Much more pronounced than other drugs
  - Neutralization of media
    - Ammonia produced by deamination of PZA
- 1. Zhang et. Al. (2002), J. Med. Microbiol. **51**(1):42-49.
- 2. Salfinger and Heifets (1988), Antimicrob. Agents Chemother. 32:1002-1004.

#### Research Plan

#### Improve MGIT testing protocol

- Hypothesis = False resistance is caused by over inoculation
- Experimental Approach
  - Test reduced inoculum sizes (i.e., # of bacilli)
  - Test Time Past Detection (TPD) of seed tube
- Pilot Project (16 strains)
  - Evaluate new dilution scheme
  - Evaluated time post positive of seed tube
  - 10 strains (with no *pncA* mutation) repeatedly found resistant using standard inoculation protocol were susceptible using a reduced inoculum

#### Research Plan

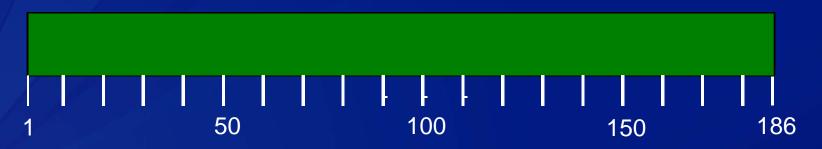
- Impact of specific pncA mutations on PZA MIC
  - Phase 1: High Concentration Range
    - PZA = 0, 300, 400, 600 & 800 μg/ml
    - NAm = 0, 1600, 1800, 2000, 2500 μg/ml
  - Phase 2: Low Concentration Range (when necessary)
    - PZA = 0, 50, 100, 200, 300 μg/ml
    - NAm = 0, 100, 300, 600, 1200 μg/ml
  - Phase 3: Susceptible Isolates (WT pncA)
    - Determine the best MIC range
    - Test new dilutions for 3-5 days post positive seed tubes

# **Molecular Diagnosis of PZA Resistance**

- What is the first requirement for development of a great molecular diagnostic?
  - A mechanism of resistance
- Mutations within pncA are associated with PZA resistance in M. tuberculosis.

#### **PncA**

- 186 amino acids
- No "hotspot" for mutations
  - DTBE LB culture collection contains isolates with mutations at 70 different amino acids
  - A database of published mutations within PncA describes isolates with mutations in an additional 45 positions
- Multiple different substitutions can occur at each position
  - For example: the histidine at amino acid 57 has been changed to an arginine, aspartic acid, leucine, proline, or tyrosine in different isolates
- Some strains contain insertions or deletions in pncA
- Some strains contain mutations in the promoter region that would likely affect the amount of PncA protein made



# Goal: Design a rapid, simple system to evaluate the effect of PncA mutations on PZase activity

### **PncA Enzymatic Assay**

- Use E. coli
- Transfer mutant M. tuberculosis pncA alleles into an E. coli strain that does not produce its own PncA
- Make expression of the mutant PncA controllable
- Examine PZase activity of mutant pncA allele
  - Add substrate to E. coli culture
  - Observe color change

# **PncA Enzymatic Assay**



Red coolor change == rfunction tabra 2 as esusee just la let

# **PncA Enzymatic Assay**

Vector Only Control



2 3

WT PncA Control



2 3 4

Tube	IPTG	PZA	<b>Expected Results</b>	
1			Resistant	
1	+	+	no color change	
2	+	-	no color change	
3	-	+	no color change	
4	-	-	no color change	

# Correlate PZA MIC with Enzymatic Activity

# pncA Mutation Collection

#### 124 strains with a SNP

- 76 with MIC and enzymatic data
  - 50 unique SNPs
  - Occur at 39 different codons

## 32 strains with an indel or promoter mutation

- 22 with MIC and enzymatic data (indels only)
  - 14 unique indels
  - 4 unique promoter mutations
  - 100% exhibited MIC > 800 μg/ml for PZA

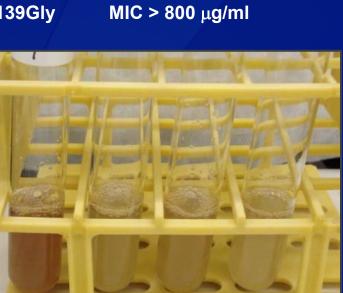
# MIC and Enzymatic Data of 50 Unique pncA nSNPs

ENZYME ACTIVITY LEVEL MIC LEVEL (µg/ml) NUMBER OF STRAINS (%)

0 (not detected)		36 (Total)
	> 800	23 (64)
	400 - 800	8 (22)
	< 300	5 (14)
1-5 (weak to very strong)		14 (Total)
	> 800	1 (7)
	400 - 800	1 (7)
	< 300	12 (86)



Val139Gly



His137Arg

MIC < 300  $\mu g/ml$ 



Asp110Gly

MIC < 300  $\mu$ g/ml



Glu37Val

MIC < 300  $\mu$ g/ml

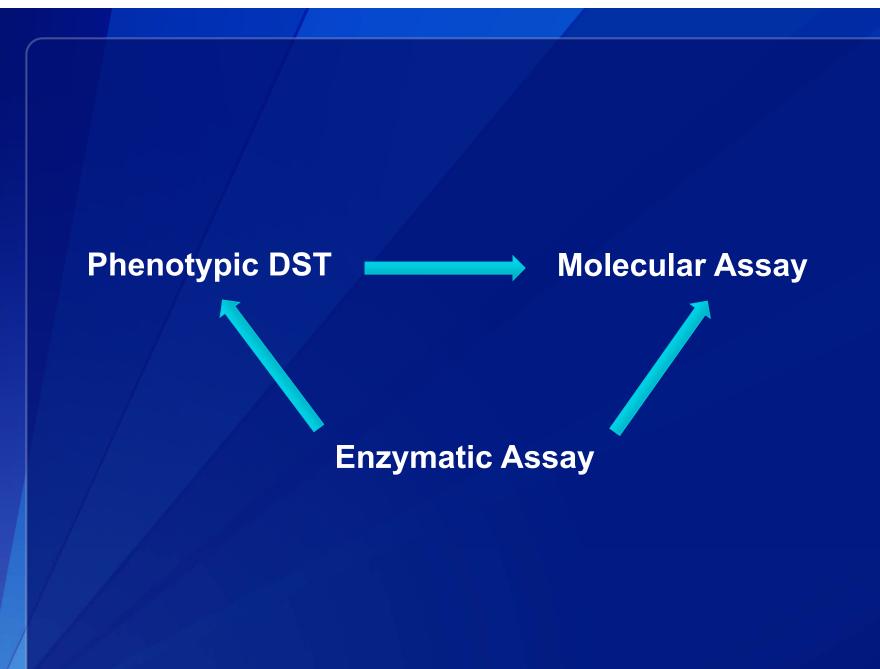
# **Interesting Mutation**

#### □ Codon 47

- Threonine to Proline (N = 2)
  - PZA MIC > 800 μg/ml
  - NAm MIC > 2500 μg/ml
- Threonine to Alanine (N = 5)
  - PZA MIC < 300 μg/ml
  - NAm MIC < 1600 μg/ml

# **Summary**

- Improved the MGIT 960 PZA assay
  - Reducing the size of the inoculum
- Developed an PncA enzymatic assay
- Correlate a pncA mutation with PZA MIC and enzyme activity
- Useful for developing molecular assays



### **Future Work**

- Continue to perform MIC and enzymatic assays
  - CDC collection
  - Collaborators
  - Susceptible strains (WT pncA)
- Structural modeling of PncA mutants
- Generate antibody to PncA
- Develop a database
  - Available to scientific community
  - MIC and enzymatic data for each mutation

# **Acknowledgments**

#### PZA TEAM

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